RHEA JANWADKAR
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Languages: English, Hindi, Marathi, Spanish

SUMMARY

Biology undergraduate (Class of 2026) specializing in Cell Biology, Molecular Biology & Genetics, with experience in synthetic biology, orthopaedic research, clinical research, and cytogenetics. Skilled in DNA cloning, histological analysis, and clinical data collection seeking an entry-level biosciences research role.

EDUCATION

Boston University, Boston, MA **Expected Graduation May 2026**

- **B.A. in Biology**, Cell Biology, Molecular Biology & Genetics specialization | Honors Research in Biology| Minor: Visual Arts | GPA: 3.73 | Dean's List
- Relevant Coursework:
  - Organic Chemistry I & II
  - Intensive Cell Biology
  - Intensive Genetics
  - Systems Physiology
  - Molecular Biology
  - Immunology
  - Carcinogenesis

RESEARCH & WORK EXPERIENCE

Honors Research in Biology, Boston University School of Medicine – Department of Orthopaedic Surgery (Sept 2025 - Present)

- Contributing to the development and analysis of a murine comminuted fracture model, performing histological assessments and collecting, recording, and analyzing experimental mouse data to investigate mechanisms of bone healing.
- Completing an Honors Research thesis, to be written and defended before a faculty committee.

Clinical Research Intern, Boston University School of Medicine – Department of Gastroenterology (Jun 2025 - Present)

- Spent 2 months observing various gastroenterology clinical trials through patient visit observation, protocol review, and monitoring activities.
- Contributing to a paper on a retrospective LI-RADS ultrasound study by assisting with clinical data collection and analysis under faculty mentorship.
- Completed required clinical research certifications, including EPIC training, Good Clinical Practice, HIPAA and Research Data Security, and Standard Operating Procedures.

Undergraduate Researcher, Wilson Wong Lab, Boston University (Sept 2022 - Dec 2024)

- Conducted wet lab research to design and modify synthetic DNA and mRNA receptor systems for applications in regenerative medicine.
- Gained proficiency in plasmid design using Benchling, DNA cloning, PCR, gel electrophoresis, and bacterial transformation, with practical experience in protein purification and transient transfection of HEK cells.

Research Intern, SDG's Genetic Centre, India (Aug 2024)

- Observed and assisted with cytogenetic procedures including slide preparation, staining, and FISH (fluorescence in situ hybridization).
- Assisted with patient reporting and gained exposure to diagnostic laboratory workflows.

Intern, Connexo Asia Limited (May 2021 - Jul 2021)

- Gained exposure through observation of pharma and biotech quality management systems, including QMS automation, software validation, and manufacturing documentation.
- Developed understanding of how quality and compliance frameworks support scalable biotech manufacturing.

TECHNICAL SKILLS

Molecular Biology: Advanced in DNA cloning, PCR, gel electrophoresis, bacterial transformation, transient transfection of mammalian cells
Animal Research: Experience in handling murine models, Von Frey assessment, histological sample analysis
Cytogenetics: Proficient in slide preparation, staining
Clinical Research: EPIC certified, clinical data collection & analysis, protocol review, patient-visit observation
Other Tools: Exposure to QMS software validation, scientific poster design, FISH

ADDITIONAL TRAINING & CERTIFICATIONS

EPIC Certification (Jul 2025)

- Certified in EPIC electronic health record systems; applied EPIC for clinical research.

Genomics: Decoding the Universal Language of Life, University of Illinois Urbana-Champaign, Coursera (Dec 2020 - Jan 2021)

- Earned course certificate and acquired foundational knowledge of genome structure, computational genomics, proteomics, epigenetics, and literature analysis on stem cell applications in genomics.